Detection of AmpC and Extended-Spectrum Beta-Lactamases in Clinical Isolates of *Pseudomonas Aeruginosa* from Patients with Cystic Fibrosis

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**ABSTRACT**

**Background and Objectives:** *Pseudomonas aeruginosa* is the most frequent opportunistic pathogen isolated from the sputum of patients with cystic fibrosis (CF). Resistance to β-lactam antibiotics may arise from over expression of the naturally occurring AmpC cephalosporinases or acquired extended-spectrum β-lactamases (ESBL). The aim of this study was to determine the antibiotic resistance profiles as well as the prevalence of ESBL and AmpC production in clinical isolates of *P. aeruginosa* from CF patients in Tehran.

**Methods:** Antibiotic resistance of 50 non-duplicate *P. aeruginosa* isolates was determined by the disc diffusion method. AmpC β-lactamase production was detected by the antagonism disc test and ESBL production was detected by the phenotypic confirmatory test. The presence of ESBL and AmpC genes was assessed by PCR, followed by sequencing the PCR products.

**Results:** The antibiotic resistance rates were as follows: 22% to ceftriaxone, 20% to cefotaxime, 10% to imipenem, 8% to carbenicillin and 6% to ticarcillin, 4% each to cefepime, tobramycin, amikacin and aztreonam and 2% to each piperacilin, meropenem and ceftazidime. AmpC production was observed in 47 isolates (94%) and ESBL production was observed in one isolate (2%). PCR results showed that all isolates carried the *bla*<sub>AmpC</sub> β-lactamase gene. One multidrug-resistant isolate carried both *bla*<sub>TEM</sub> and *bla*<sub>PER-1</sub> genes.

**Conclusion:** The results showed that despite the low rate of antibiotic resistance in *P. aeruginosa* CF isolates, the presence of multiple β-lactamases even in one isolate is alarming and can complicate the already difficult treatment of chronic infections in the lungs of CF patients.

**KEYWORDS:** *Pseudomonal aeruginosa; Cystic Fibrosis; ESBL, AmpC, Beta-Lactamases.*
INTRODUCTION

Chronic Pseudomonas aeruginosa lung infection is the main cause of morbidity and mortality in cystic fibrosis (CF) patients. About 80% of adults with CF suffer from the chronic P. aeruginosa infection (1,2). In the lung of CF patients, P. aeruginosa exhibits several genotypic and phenotypic changes such as antibiotic resistance and mucoid conversion (3). P. aeruginosa uses a wide variety of different resistance mechanisms and is essentially resistant to all classes of commonly used antibiotics (4). Several mechanisms are responsible for resistance to β-lactam antibiotics in P. aeruginosa including over expression of chromosomally mediated AmpC cephalosporinases, acquisition of transferable β-lactamase genes such as extended-spectrum β-lactamases (ESBLs) and metallo-β-lactamases (MBLs), presence of efflux systems, and reduced membrane permeability (5,6). In P. aeruginosa, the decreased susceptibility to the extended-spectrum cephalosporins such as ceftazidime mostly results from over expression of the naturally occurring AmpC β-lactamases (6). ESBLs are capable of hydrolyzing penicillins, cephalosporins and aztreonam (except for cephapemycin or carbopenems). These enzymes are inhibited by β-lactamase inhibitors such as clavulanic acid (7). Up to 32 different ESBLs have been detected in P. aeruginosa belonging to class A (TEM, SHV, CTX-M, PER, VEB, GES, BEL), and class D (OXA type) β-lactamases (8). We studied the antibiotic resistance profiles in clinical isolates of P. aeruginosa from CF patients, by phenotypic detection of AmpC and ESBL production and their comparison in the presence of AmpC, TEM, SHV, CTX-M, PER and VEB β-lactamase genes among these isolates.

MATERIAL AND METHODS

Fifty non-duplicate clinical isolates of P. aeruginosa were collected from CF patients at Mofid Children’s Hospital in Tehran between July 2012 and February 2013. Twenty-nine (58%) isolates were obtained from males and 21 (42%) were obtained from female patients (age range, 6 months to 26 years). The mean age (± SD) was 92.52 (± 70.99) months with a median of 72 months. The isolates were identified by Gram staining and conventional microbiological tests including oxidase production, blue–green pigmentation on Mueller Hinton agar [MHA, (Liofilchem, Italy)], acid and gas production on triple sugar iron agar slants (BBL, USA), and growth at 42°C (9). The isolates were maintained in brain heart infusion broth (Oxoid, UK) containing 10% dimethyl sulfoxide at -20 °C until use. P. aeruginosa PA01 containing bla_aper (kindly provided by Dr Abdi, Alzahra University, Iran), DNA from K. pneumoniae 7881 harboring blatem, blashv and blactxm, P. aeruginosa KOAS carrying blasPER and P. aeruginosa 10.2 harboring blavesBL (obtained from Pasteur Institute, Tehran, Iran) were used as controls for PCR experiment. P. aeruginosa ATCC 27853 was used as control for susceptibility tests. Antibiotic susceptibility testing of the isolates was carried out by the disc diffusion method according to the Clinical and Laboratory Standard Institute (CLSI) guidelines (10). The antibiotic discs (MAST Group LTD, Merseyside, UK) used in this study included: ceftazidime (30 μg), cefotaxime (30 μg), ceftriaxone (30 μg), cepefime (30 μg), cefoxitin (30 μg), imipenem (10 μg), meropenem (10 μg), carbencillin (100 μg), ticarcillin (75 μg), piperacillin (100 μg), amikacin (30 μg), tobramycin (10 μg) and aztreonam (30 μg). Minimum inhibitory concentrations (MICs) for imipenem and ceftazidime were determined by the microdilution assay as recommended by the CLSI (10). Initial screening for AmpC β-lactamase production was performed by the disc antagonism test on the isolates showing a zone diameter of <18 mm to cefoxitin disc (11). Briefly, a lawn culture of each test isolate (0.5 Mc Farland) was prepared on a MHA plate, and then ceftazidime (30μg) and cefoxitin (30μg) discs were placed 20 mm (centre to centre) apart from each other followed by incubation at 37 °C for 18-20 hours. Inducible AmpC β-lactamase production was detected when blunting of ceftazidime inhibition zone occurred adjacent to the cefoxitin disc. ESBL production was detected by the conventional double disc synergy test using ceftazidime (30 μg) and cefotaxime discs (30 μg) with or without clavulanic acid (10 μg) as recommended by the CLSI (10). An increase of ≥ 5 mm in the inhibition zone of either cephalosporin in combination with clavulanic acid compared to the cephalosporin alone was interpreted as ESBL production. DNA extraction was performed by boiling method. Briefly, a
loopful of an overnight grown culture of each isolate was suspended in 500 μl of sterile double distilled water, boiled at 100 °C for 10 min and then centrifuged at 12000 g for 10 min. The supernatant was used as DNA template for PCR amplification tests. PCR amplifications were performed using specific primers for Ambler class A (blaTEM, blaSHV,blaCTX-M, blaVEB-1, blaPER-1) and class C (blaAmpC) β-lactamase genes (Table 1). PCR experiments were carried out in 25 μl volume reaction mixtures containing 10 pM of each primer, 200 μM dNTP, 1.4 mM MgCl2, 1 μl of DNA template and 1 U of Taq polymerase in the reaction buffer provided by the manufacturer (CinnaGen, Tehran, Iran). PCR process was carried out in a thermocycler (Technne, UK) using the following program: 5 min denaturation at 95 °C, followed by 30 cycles at 95 °C for 1 min, 1 min at the annealing temperature (43-63°C for ESBLs and 62°C for AmpC), 1 min at 72 °C and final extension for 10 min at 72 °C.

Table1- Primers used for PCR detection of ESBLs and AmpC genes

<table>
<thead>
<tr>
<th>β-lactamase</th>
<th>Primer</th>
<th>Sequence (5' to 3')</th>
<th>Product size (bp)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Class A</td>
<td>TEM-F</td>
<td>GAGTATCTACGATTTGCGTC</td>
<td>851</td>
<td>12</td>
</tr>
<tr>
<td></td>
<td>TEM-R</td>
<td>ATATGACGATGCGGCCATACCT</td>
<td>231</td>
<td>12</td>
</tr>
<tr>
<td></td>
<td>SHV-F</td>
<td>AGAGATCCACTATCGGCCAGCAG</td>
<td>550</td>
<td>13</td>
</tr>
<tr>
<td></td>
<td>SHV-R</td>
<td>ATTCAGTTCCGTTCGCGGCGG</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>CTX-M-F</td>
<td>CGCTTTGCCAGGTGAGCACT</td>
<td>500</td>
<td>13</td>
</tr>
<tr>
<td></td>
<td>CTX-M-R</td>
<td>ACCCGGATATGCTTGCTGGT</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>PER-1-F</td>
<td>ATGAAATGCATTATGAAAGGC</td>
<td>920</td>
<td>12</td>
</tr>
<tr>
<td></td>
<td>PER-1-R</td>
<td>AATTGTCGCTGCTAGGCGAAGA</td>
<td>643</td>
<td>12</td>
</tr>
<tr>
<td></td>
<td>VEB-1-F</td>
<td>CGACTTTCAATTCGCGATGC</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>VEB-1-R</td>
<td>GGACTCTGCAACAAATACGCC</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Class C</td>
<td>AmpC-F</td>
<td>ATGCAGCGCATAGCCGCGCCTC</td>
<td>1243</td>
<td>14</td>
</tr>
<tr>
<td></td>
<td>AmpC-R</td>
<td>CGCCCTGCGCCGCGCCTTC</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

RESULTS

Among the 50 CF isolates, 48 (96%) were resistant to cefoxitin, one was susceptible, and one showed intermediate resistance. Resistance rates to other antibiotics were as follows (Figure 1): ceftriaxone (22%), cefotaxime (20%), imipenem (10%), carbenicillin (8%), ticarcillin (6%), cefepime, amikacin, tobramycin and aztreonam (4%) and piperacilin, ceftazidime and meropenem (2%). Overall, the highest resistance rate was to ertapenem and the lowest was to piperacilin, ceftazidime and meropenem. One isolate (2%) was multidrug-resistant with MIC ≥16 μg/ml for imipenem and ≥128 μg/ml for ceftazidime.

Figure 1–Antibiotic resistance profiles of the 50 P. aeruginosa CF isolates, CAZ, ceftazidime; CTX, cefotaxime; CRO, ceftriaxone; CPM, cefepime; IPM, imipenem; MEM, meropenem; PY, carbenicillin; TC, ticarcillin; PRL, piperacilin; AK, amikacin; TN, tobramycin; ATM, aztreonam.
AmpC production was observed in 47 isolates (94%) and ESBL phenotype was detected in one isolate (2%). PCR amplification results showed that all isolates carried the bla\textsubscript{AmpC} β-lactamase gene and the multidrug-resistant isolate carried bla\textsubscript{AmpC}, bla\textsubscript{TEM} and bla\textsubscript{PER-1} genes (Figure 2). bla\textsubscript{PER}, bla\textsubscript{CTX-M} and bla\textsubscript{VEB} genes were not detected in any of the isolates. Sequence analysis of the PCR products showed the presence of a new allele for the bla\textsubscript{TEM} gene (www.lahey.org/studies/temtable.asp) (15). Overall, there was a good correlation between β-lactamase phenotype and its related gene carriage among the tested isolates (p<0.05).

DISCUSSION

β-lactam antibiotics are often inefficient in the treatment of \textit{P. aeruginosa} lung infections. The main reasons for treatment failure are thought to be ESBL production, hyperproduction of chromosomally encoded AmpC β-lactamases and biofilm formation (1). The current CLSI guidelines have not provided a method for detection of AmpC β-lactamases. It has been suggested that cefoxitin-resistant isolates (showing a zone diameter of <18 mm to cefoxitin disc) are potential AmpC β-lactamase producers and the disc antagonism test should be used for screening AmpC β-lactamase production in these isolates (11,16). In this study, all 50 CF isolates carried the bla\textsubscript{AmpC} gene, while 47 isolates (94%) were AmpC producers. These results show a good correlation between AmpC gene carriage and AmpC phenotype among the isolates. Of the three bla\textsubscript{AmpC} positive and AmpC negative isolates, one was susceptible to all tested antibiotics, two were resistant to cefoxitin, and one also showed intermediate resistance to cefotaxime. However, the susceptibility of these isolates to other 3\textsuperscript{rd} generation cephalosporins was not investigated in the present study. The presence of genes does not necessarily mean that they are expressed, which could be influenced by the environment as well as other regulatory mechanisms. Limited number of studies on AmpC production by \textit{P. aeruginosa} CF isolates are available. In a study in Africa, AmpC production was not detected in any of the 15 \textit{P. aeruginosa} isolates from CF patients (17).

The rate of ESBL production in \textit{P. aeruginosa} CF isolates varies worldwide. In a study in Iran, Vali et al. showed that 9.6% of 52 \textit{P. aeruginosa} CF isolates carried bla\textsubscript{CTX-M} (18). Another study in Iran detected the bla\textsubscript{PER-1} gene in 10% of \textit{P. aeruginosa} CF isolates (19). Mhlongo et al. reported the ESBL phenotype in all of their tested isolates (17). Other studies have reported the presence of bla\textsubscript{CTX-M} gene and coproduction of PER-1 and VIM-2 in one \textit{P. aeruginosa} CF isolate (20, 21). We found ESBL production in one multidrug-resistant isolate, which also carried bla\textsubscript{AmpC}, bla\textsubscript{TEM} and bla\textsubscript{PER-1} genes. Overall, the spread of different ESBL genes in \textit{P. aeruginosa} CF isolates is increasing. The discrepancy between β-lactamase phenotype and gene carriage could be due to environmental conditions such as the presence of antibiotics that may allow the expression of β-lactamase genes in vivo but not in vitro.

CONCLUSION

The results showed that despite the low rate of antibiotic resistance in \textit{P. aeruginosa} CF Isolates, the presence of β-lactamase genes could lead to their
expression under selective environmental pressures such as extensive use of antibiotics, resulting in treatment failure in these infections. In addition, resistance genes from even one multidrug-resistant isolate can spread among the susceptible strains and cause increased morbidity and mortality among the CF patients.

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CONFLICT OF INTEREST:
There are no conflicts of interest.

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