**Antibacterial Activity of Aqueous and Methanolic Extracts of *Crocus sativus* Stigma and *Cinnamomum cassia* against Clinical Isolates of some Gram-Positive and Gram-Negative Pathogenic Bacteria**

**Background and Objectives:** Medicinal and aromatic plants are sources of natural antimicrobial compounds that could be useful replacements for antibiotics. The aim of this study was to assess antimicrobial activity of *Crocus sativus* stigma and *Cinnamomum cassia* extracts against some Gram-positive and Gram-negative bacteria.

**Methods:** Antimicrobial activity of methanolic and aqueous extracts of the plants was tested against clinical isolates of *Klebsiella pneumonia*, *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Escherichia coli* and *Enterococcus* using the microdilution method. Minimal inhibitory concentration (MIC) and minimum bactericidal concentration of each extract against the mentioned bacteria were also determined.

**Results:** The MIC of the methanolic extract of *C. cassia* was 80 µg/ml against *Enterococcus*, *K. pneumonia* and *E. coli*. The MIC of the methanolic extract of *C. sativus* was 160 µg/ml against *Enterococcus* and *S. aureus*. The minimum bactericidal concentration of the methanolic extracts of *C. sativus* and *C. cassia* was 320 µg/ml against *K. pneumonia* and 160 µg/ml against *Enterococcus*.

**Conclusion:** The extracts of *C. sativus* and *C. cassia* exhibit promising antibacterial activities against clinical isolates of the tested bacteria. Our results suggest that the extract of these plants can be further exploited as potential antibacterial agents against multi-drug resistant bacteria.

**Keywords:** *Cinnamomum* aromaticum, Crocus, Anti-Bacterial Agents.

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Najmeh Jomehpour (PhD)  
Department of Microbiology and Virology, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran

Kiarash Ghazvini (PhD)  
Antimicrobial Resistance Research Center, Baali Research Institute, Mashhad University of Medical Sciences, Mashhad, Iran, and Department of Microbiology and Virology, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran

Mahshid Jomehpour (BSc)  
Lab Sciences, Shahrood University of Medical Sciences

**Corresponding author:** Najmeh Jomehpour  
Email: Jomehpourn961@mum.ac.ir  
Tel: +98-9153314525  
Address: Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran

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INTRODUCTION

Infections caused by pathogenic bacteria have been considered a main cause of morbidity and mortality in humans (1), particularly following the emergence and spread of multidrug-resistant bacteria which have increased the risk of treatment failure (2). Hence, it is crucial to find alternative antibacterial agents with therapeutic potential against antibiotic-resistant pathogens. The discovery and use of medicinal plants go back to prehistoric times (3). It is well-demonstrated that extract of some medicinal plants possess antibacterial, antimitogenic, antithrombotic and vasodilatory properties (4). Therefore, medicinal plants are currently investigated as a major source of natural pharmaceutically valuable drugs that could be used for treatment of human diseases (5).

*Crocus sativus*, commonly known as saffron crocus, is a plant species from the family **Iridaceae** that is native to Southern Europe but currently cultivated worldwide, predominantly in Iran, Spain, Italy and Greece (6). Saffron is the stigma of the plant, which has been used for treatment of several medical conditions, such as gastrointestinal disorders, urinary tract infections, catarrhal and malignancies (7). Cinnamon is another well-known medicinal plant that belongs to the genus *Cinnamomum*, family **Lauraceae**, which is widely distributed in China, India and Australia (8). *Cinnamomum cassia* has been shown to have choleretic, anti-ulcer, antifungal, antibacterial, antiviral and anti-inflammatory properties (9). *Cinnamomum* bark is rich in cinnamaldehyde, a highly electronnegative compound that inhibits growth of microorganisms by interfering in biological processes involving electron transfer and interacting with nitrogen-containing components, e.g. proteins and nucleic acids (10). The aim of this study was to assess antimicrobial activity of *C. sativus* stigma and *C. cassia* extracts against some Gram-positive and Gram-negative bacteria.

MATERIALS AND METHODS

*C. sativus* (stigma) and *C. cassia* were obtained from a local herbal market in Torbat-e Heydariyeh and Mashhad, Iran. The plants were shade dried at room temperature. Dried plants were ground to fine powder using a homogenizer. Methanolic and aqueous extracts of the plants were obtained by soaking 5 g of plant powder into 50 mL of denatured MeOH (95%) and sterile distilled water in a flask for 72 hours, respectively. The flasks were agitated daily. Stock concentrations of 100 mg/mL of dry extracts in dimethyl sulfoxide were prepared, sterile filtered (0.2 μm) and stored in darkness at 25 °C.

Clinical isolates of Gram-positive (Staphylococcus aureus, Enterococcus and Streptococcus pyogenes) and Gram-negative (Escherichia coli, Klebsiella pneumonia and Pseudomonas aeruginosa) bacteria were obtained from patients hospitalized in the Ghaem hospital, Mashhad, Iran. The bacteria were grown overnight in brain heart infusion (BHI) broth (LioFilchem, Italy) at 37 °C. After obtaining a turbidity equivalent to that of a 0.5 McFarland standard (10^6 CFU/mL), bacterial suspensions were adjusted with sterile saline to a concentration of 1 × 10^7 CFU/mL.

Minimum inhibitory concentration (MIC) of the extracts against the bacteria was determined using the microdilution method as recommended by the Clinical and Laboratory Standards Institute (CLSI) (11). The MIC was defined as the lowest concentration of extract that inhibited growth of bacteria (12). The stock solutions of both extracts were diluted and transferred into first well of a 96 well plate, and serial dilutions were made to obtain concentration range of 80–320 μg/ml. The plate was incubated at 37 °C for 24 hours in aerobic conditions. Then, 200 μl of tetrazolium chloride solution (0.5%) were added to all wells. The plates were examined for color change after 3 hours. To confirm the MIC and minimum bactericidal concentration (MBC), 50 μl were taken from wells with no visible growth and inoculated in Müller-Hinton agar. After 24 hours of aerobic incubation at 37 °C, the numbers of viable microorganisms was determined.

The MBC was then recorded as the lowest concentration that killed at least 99.99% of bacteria (13). All experiments were repeated at least three times. The same concentrations of ethanol and bacteria without the extracts were used as the negative and positive controls, respectively.
RESULTS

Table 1 presents the MIC and MBC values of the methanolic and aqueous extract of C. sativus stigma against the tested bacteria. The MIC of the methanolic extract of C. sativus stigma was 320 μg/ml against P. aeruginosa, S. pyogenes, E. coli and K. pneumonia and 160 μg/ml against S. aureus and Enterococcus. The MBC of this extract was 160 μg/ml against Enterococcus and 320 μg/ml against other bacteria. The MIC of the aqueous extract of C. sativus was 320 μg/ml or higher against all tested bacteria except for E. coli (160 μg/ml).

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<tr>
<th>Microorganism</th>
<th>Methanolic extract</th>
<th>Aqueous extract</th>
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<tr>
<td></td>
<td>MIC (μg/ml)</td>
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<td>P. aeruginosa</td>
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<td>S. pyogenes</td>
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<td>S. aureus</td>
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Table 2 shows the MIC and MBC values of the methanolic and aqueous extracts of C. cassia against the tested bacteria. The MIC of the methanolic extract of C. cassia was 80 μg/ml against Enterococcus, K. pneumonia and E. coli and 160 μg/ml against P. aeruginosa, S. pyogenes and S. aureus. According to the results, P. aeruginosa, Enterococcus, S. pyogenes, and S. aureus were resistant to the aqueous extract of C. cassia (Table 2).

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<td>S. aureus</td>
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DISCUSSION

Extensive antibiotic use over the years has led to the emergence and spread of antibiotic-resistant bacterial strains. Plant-derived antimicrobial compounds with antibacterial properties could be suitable alternatives to antibiotics for treatment of infections caused by antibiotic-resistant bacteria. In this study, we examined the inhibitory activity of C. sativus stigma and C. cassia extracts against some Gram-positive and Gram-negative bacteria. The results showed that both plant extracts exhibit varying degree of inhibitory effects on the clinically isolated bacteria. The MIC of the methanolic and aqueous extracts of C. sativus stigma against E. coli was 320 μg/ml and 160 μg/ml, respectively. In a previous study, the MIC of C. sativus stigma extract against P. aeruginosa and E. coli was 300 μg/ml and 500 μg/ml, respectively. In addition, the MIC and MBC of the extract against S. aureus was 200 μg/ml and 300 μg/ml, respectively (14). However, in our study, the MIC of the methanolic and aqueous extracts of C. sativus stigma against S. aureus was 160 μg/ml and 320 μg/ml, respectively.

In a study by Pintado et al., the MIC of safranal against S. aureus and E. coli was 4 mg/ml (15). Methanolic extracts of various Crocus spp. were found to have significant inhibitory activity against different bacteria (16). The antimicrobial properties of saffron extracts are mainly attributed to safranal and crocin (17), while cinnamaldehyde is the main active antibacterial constituent of cinnamon (18). In our study, the MIC of the methanolic extract of C. cassia was 160 μg/ml against P. aeruginosa. Inconsistent with this finding, Gupta et al. claimed that P. aeruginosa was resistant to C. cassia extract. The MIC values reported by Gupta et al. for S. aureus (62.5 mg/ml) and E. coli (1000 mg/ml) were also
significantly higher than the values we found in the present study (18). In another study, the ethanolic extracts of cinnamon and turmeric effectively inhibited growth of *Bacillus subtilis* and *E. coli*, which implies that the extracts could be equally effective against both Gram-negative and Gram-positive bacteria (19).

In our study, the methanolic extract of *C. sativus* was more effective than the aqueous extract against growth of *S. pyogenes* and *S. aureus*. Given the results of the MIC assay, it can be concluded that the methanolic extract of *C. cassia* was more effective than the methanolic extract of *C. sativus* against *Enterococcus*, *K. pneumonia* and *E. coli*.

**CONCLUSION**

The extracts of *C. sativus* and *C. cassia* exhibit promising antibacterial activities against clinical isolates of *S. aureus*, *E. coli*, *P. aeruginosa*, *Enterococcus*, *S. pyogenes* and *K. pneumoniae*. Based on the results of the MIC assay, it can be inferred that the methanolic extracts are generally more effective than the aqueous extract against the tested pathogens. Our results suggest that the extract of these plants can be further exploited as potential antibacterial agents against multidrug-resistant bacteria.

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**CONFLICT OF INTEREST**

All authors declare that there is no conflict of interest regarding the publication of this article.


