Background and objectives: Tricyclazole (TZ) is a member of triazole fungicides, which might cause damage in living systems. This study was carried out to examine effects of TZ on liver tissues and level of liver enzymes.

Methods: Forty mice were randomly divided into four groups including control, sham and two experimental groups. Experimental groups 1 and 2 received 5 mg/Kg and 15 mg/Kg intraperitoneal injection of TZ for two weeks, respectively. The sham group received sterile water but the control group received no injection. The animals were sacrificed 24 h after the last injection, and microscopic slides were prepared for cell counting and evaluation of tissue damage. Levels of liver enzymes were measured using commercial kits. Data was analyzed in SPSS (version 20) using one-way ANOVA.

Results: The injection of TZ caused a significant increase in the number of hepatocytes and a significant decrease in the number of Kupffer cells compared to control group (P<0.001). In the experimental group, the level of alanine aminotransferase and aspartate aminotransferase increased, but the level of alkaline phosphatase decreased significantly compared to control group (P<0.001). We also detected several forms of tissue damage including necrosis and degeneration of hepatocytes, hyperplasia, and penetration of inflammatory cells and expansion of sinusoids.

Conclusion: Our results indicate that the intraperitoneal injection of TZ in mice can cause irreparable hepatic damage in a dose-dependent manner.

Keywords: Tricyclazole, hepatocytes, Alanine, Aspartate aminotransferase.
INTRODUCTION

Widespread application of toxins can cause teratogenic effects on various growth-related phenomena, which has raised concerns about the potential damages that pesticides can inflict on human health and the ecosystem (1). Tricyclazole (TCZ) is one of the most commonly used pesticides, particularly in Asian countries. This systemic fungicide belongs to the triazole group, and used to eradicate rice blast (2, 3). This toxin can remain active in soil and water after long-term use. Exposure to the toxin can cause several health-related problems (4). Toxins are readily absorbed by skin and converted to secondary metabolites in the liver. A large amount of these metabolites are excreted by the kidneys, but the remaining may cause tissue damage in the body (5). Toxicity of TCZ in various organs including the reproductive system and liver and the potential biochemical and tissue changes have been reported (6, 7). Some reports indicate that triazoles can affect liver enzymes, particularly cytochromes. Although the precise mechanism of action of TCZ in the body has not been determined, some scientists believe that this toxin promotes lipid peroxidation and cell death by inducing production of free radicals and reactive oxygen species (8). Studies on liver tissues of prenatal mice showed that triazoles increase the number of hepatocytes and concentration of alanine transaminase (ALT) and aspartate aminotransferase (AST), but reduce the activity of alkaline phosphatase (ALP). In addition, some studies demonstrated that TCZ significantly increases the activity of ALP and ALT in fish (9, 10). As the largest gland in the body, liver has several important functions including neutralization of toxins, drugs and other harmful compounds. Detoxification and metabolic activation by liver enzymes usually produce toxic metabolites that cause toxic damage, necrosis and acute liver failure (11). Despite the widespread use of TCZ in recent years, little information is available about acute hepatotoxicity in Iran. Therefore, the present study has examined effects of TCZ on histopathology of liver and level of liver enzymes.

MATERIAL AND METHODS

In this experimental-analytical study, we purchased 40 adult NMRI mice aged two months (average weight 30 ± 5 g) from the Pasteur Institute (experimental Animal keeping center, Iran). The animals were kept in special cages at 23 ± 2 °C and 12:12 h light/dark cycle. The animals were given ad libitum access to food and water. The study was in accordance with ethical standards for handling animals set by the Ethics Committee of Islamic Azad University (No: 7812). From the stock solution of TCZ 95% (Gol Sam Co, Gorgan, Iran), various concentrations of the toxin were prepared in sterile water and then injected to the animals on a daily basis and at a specific time. The animals were randomly divided into four equal groups (n=10). Control group received no TCZ and sham group received sterile water injections for two weeks. Experimental group 1 received 5 mg/Kg intraperitoneal injection of TCZ for two weeks (5 days a week, while resting for two days). Experimental group 2 received 15 mg/Kg intraperitoneal injection of TCZ for two weeks (5 days a week, while resting for two days)[12]. All animals were kept under optimum conditions. Twenty-four hours after receiving the last injection, the animals were sacrificed and sampling was done. Blood samples were taken from heart to measure levels of ALT, AST and ALP. The samples were then centrifuged at 3000 rpm for 15 min to separate serum. Next, the level of each enzyme was measured using Hitachi 911(Taiwan) full automated analyzer and Pars Azmoon kits (Iran). The mice were dissected and liver tissues were isolated. The tissues were fixed in 10% formalin. After routine steps of tissue preparation, paraffinized tissue blocks were retrieved and used to prepare 5-micron serial sections stained with hematoxylin-eosin. The sections were selected in a way that almost 100 sections from each part of the liver could be evaluated. Cell counting for hepatocytes and Kupffer cells was carried out with three repeats using eyepiece graticules. The data was analyzed in SPSS (version 20) using one-way ANOVA and post-hoc Tukey to compare means between the groups. P-values less than 0.05 were considered as statistically significant.

RESULTS

The number of hepatocytes in the experimental groups reduced significantly compared to the control group, but the number of Kupffer cells increased significantly in the experimental
Experimental group 1 showed signs of necrosis, degeneration, and irregular hepatocyte layers, penetration of inflammatory cells, liver fibrosis and expansion of sinusoids when compared to controls. In experimental group 2, liver damages were more extensive including hepatocytes necrosis and degeneration, triad fibrosis, necrosis of parenchymal liver cells and bile duct hyperplasia. The number of Kupffer cells in experimental group 2 was higher than that in experimental group 1, which could be due to the higher dose of TCZ administered (Figure 1).

Although TCZ is converted to active metabolites in the liver, the activity of liver against these compounds is limited. Thus, exposure to large amounts of this toxin may cause toxic damage, necrosis and acute liver failures (Experimental group 1) showed signs of necrosis, degeneration, and irregular hepatocyte layers, penetration of inflammatory cells, liver fibrosis and expansion of sinusoids when compared to controls. In experimental group 2, liver damages were more extensive including hepatocytes necrosis and degeneration, triad fibrosis, necrosis of parenchymal liver cells and bile duct hyperplasia. The number of Kupffer cells in experimental group 2 was higher than that in experimental group 1, which could be due to the higher dose of TCZ administered (Figure 1).

Table 1- Mean levels of liver enzymes and the number of hepatocytes and Kupffer cells in each study group

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Control group</th>
<th>Sham group</th>
<th>Experimental group 1 (5mg/kg)</th>
<th>Experimental group 2 (15mg/kg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number of hepatocyte</td>
<td>57.6±4.6c</td>
<td>56.3±4.2a</td>
<td>49.3±3.4b</td>
<td>45.8±5.4d</td>
</tr>
<tr>
<td>Number of Kupffer cells</td>
<td>5.38±2.45a</td>
<td>5.66±2.87c</td>
<td>8.26±3.08b</td>
<td>9.73±2.96d</td>
</tr>
<tr>
<td>AST(IU/L)</td>
<td>165.5±18.4a</td>
<td>172.9±22a</td>
<td>435±25c</td>
<td>526±24c</td>
</tr>
<tr>
<td>ALT(IU/L)</td>
<td>73±11a</td>
<td>82±19a</td>
<td>152±17b</td>
<td>262±26c</td>
</tr>
<tr>
<td>ALP(IU/L)</td>
<td>323±31.4a</td>
<td>315±28.3c</td>
<td>219.6±38.36c</td>
<td>193.5±31c</td>
</tr>
</tbody>
</table>

Data shown as mean±SE. In each row, there are mean values that have at least one letter in common; these mean values are not significantly different at 5% in Duncan test.

DISCUSSION

In our study, TCZ injection reduced the number of hepatocytes but increased the number of Kupffer cells compared with controls. The level of ALT and AST in TCZ-treated groups increased significantly compared with controls. However, the level of ALP in the experimental groups reduced significantly. TCZ injection also caused damages including necrosis, hepatocyte degeneration, triad fibrosis, bile duct hyperplasia, penetration of mononuclear cells, and sinusoid expansion. These effects were more severe in mice treated with higher doses of TCZ.

Although TCZ is converted to active metabolites in the liver, the activity of liver against these compounds is limited. Thus, exposure to large amounts of this toxin may cause toxic damage, necrosis and acute liver failures.
The Effects of TCZ on Hepatotoxicity

The histopathological changes in liver tissue following TCZ administration can be an important sign of liver degeneration, which in turn increases the release of AST and ALT. The number of Kupffer cells increased after TCZ injection. In fact, these cells are mature macrophages in liver sinusoids, and the increased number of Kupffer cells for digestion of necrotic cells after TCZ injection could be attributed to their phagocytotic role.

CONCLUSION

Our results indicate that the intraperitoneal injection of TCZ in mice can cause irreparable hepatic damage in a dose-dependent manner. Considering the widespread use of TCZ as fungicide in farms and the results obtained in our study, it is recommended to accurately monitor and control the amount and duration of the toxin’s application.

ACKNOWLEDGEMENTS

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CONFLICT OF INTEREST

None declared.

REFERENCES

8. Bagchi D, Bagchi M, Hassoun EA, Stohs SJ. In vitro and in vivo generation of reactive oxygen species, DNA damage and lactate dehydrogenase leakage by selected pesticides. Toxicology. 1995; 104(1-3): 129-140.